

ISSN : 2321-9602



Indo-American Journal of Agricultural and Veterinary Sciences



editor@iajavs.com
iajavs.editor@gmail.com



Wheat grain microfungi in the Polissya region

Sumalatha T¹, Shyamala²

Abstract

Quantitative and qualitative information on the micromycetes found in Polissya-grown wheat grain is presented in this article. Wheat grain samples from the Polissya area had an average of 2.88 10⁴ 3.62 10³ colony forming units (CFU) per 1 g of grain throughout the investigation period. Twenty fungal species from nine different taxa were found in Polissya wheat kernels. The samples were infected with fungi from the genera *Alternaria* (92.5%), *Mucor* (92.5%), *Aspergillus* (83.1%), *Penicillium* (47.2%), *Fusarium* (60.4%), *Phoma* (15.1%), *Mycelia* (15.1%), *Trichotecium* (1.9%), and *Monascus* (1.9%). Among the *Aspergillus* species, the detection rates were lowest for *Aspergillus niger* (17.0%), *Aspergillus candidus* (9.4%), and *Aspergillus terreus* (1.9%). Species of *Fusarium* were found in 17.0% of the samples; they included *Fusarium oxysporum*, *Fusarium moniliforme*, *Fusarium semitectum* in (5.7%), and *Fusarium culmorum* (3.8%). The wheat grain's endophytic mycobiota consists of a very modest number of species of tiny fungus. Pure cultures were produced from *F. sporotrichiella* var. *tricinctum* isolate 1218/4 and *F. sporotrichiella* isolate 1218/5, both of which were isolated from the Polissya zone. The test culture of *Candida pseudo-tropicalis* strain 44 PC was resistant to the mycotoxins produced by these isolates, while the growth of *F. sporotrichiella* var. *tricinctum* 1218/4 was inhibited in a zone with a relative toxicity (Rf) of 0.05. This isolate also generated an unknown trichothecene mycotoxin (TTMT). *Aspergillus flavus* isolate 1219/3 was the first to create kojic and aspergillic acids, while *Aspergillus flavus* isolate 1221/1 was the second to synthesis penicillic and aspergillic acids, respectively, among the isolated fungus. Chickens of the meat and egg breed Adler Silver were given the sorbent "Mikosorb" to lessen the harmful effects of the mycotoxin deoxynivalenol on their bodies. Deoxynivalenol's toxic effects on the bodies of hens in the experimental group were found to be mitigated when the sorbent "Mikosorb" was added to the diet at a rate of 2.0% of the total feed weight. The 12.0% drop in avian deaths provided more confirmation of this.

Keywords: Microscopic fungi; wheat grain; mycotoxins; deoxynivalenol; sorbent "Mycosorb"; Adler silver cross chickens.

1. Introduction

Wheat is one of the main cereal grains used as human food and animal feed. The problem of contamination of wheat grain with microscopic fungi, and as a result, their secondary metabolites, is one of the main factors determining the health of animals and humans. Therefore, it is essential to study the contamination of wheat crops with microscopic fungi, especially pests, and diseases, in the case of long-term storage. Using various scientific approaches to improve grain quality, studying fungi that infect seeds during storage and grain cultivation and processing is a crucial task today (Stuper-Szablewska & Perkowski, 2014; Jangol, 2018; Jaroshenko et al., 2018; Minooeianhaghighi et al., 2021). Micromycetes that infect wheat seeds are often capable of synthesizing toxic metabolites that cause poisoning in humans

and animals and have carcinogenic and cumulative properties. To date, more than 250 microscopic fungi are known that can produce up to 500 secondary metabolites of different chemical nature, which are united by the common name "mycotoxins" (Vasjanovych et al., 2016; 2017; Iliff et al., 2022). The root causes of the increase in the distribution and harmfulness of toxin-forming micromycete species are several factors, among which the main ones are the change in the phytopathological situation in agroecosystems due to long-term systematic violation of the requirements of farming systems, as well as highly favorable weather conditions for the development of micromycetes that have developed over the past 5–10 years (Tymoshchuk et al., 2014).

1. Assistant professor, Department of Pharmaceutical Analysis, Balaji College of Pharmacy, Vishakapatnam
2. Assistant professor, Department of Pharmacology, Balaji College of pharmacy, Etcherla, Srikakulam.



Micromycetes are potential phytopathogenic agents that can infect plants both during the growing season and during storage of grain and grain fodder, reducing its nutritional value and, under appropriate conditions, accumulating mycotoxins in it (Rozhkova et al., 2017; Felšöciová et al., 2021; Kolomiets et al., 2022).

The study of wheat grain micromycetes is devoted to the work of various scientists: (Hmel'nyc'kyj et al., 2012; Kotyk et al., 2013). The issues of mycotoxin content in cereals were studied by (Kotsyumbas et al., 2010; Dvorska, 2013).

The contamination of food and feed with mycotoxins is currently being studied worldwide. Mycotoxins in grain are currently detected in Europe, the USA, Africa, Asia, and Australia. Toxicological studies (Antonjak et al., 2010; Tsvilikhovskiy et al., 2010) have shown that almost 25–40 % of grain is contaminated with mycotoxins annually, and losses caused by fungal contamination can reach tens of billions of dollars per year (Golovchak, 2007; Voloshchuk et al., 2017). Most of the mycotoxins produced by *Fusarium* fungi are derivatives of 12,13-epoxytrichothec-9-ene. The chemical structure of trichothecene mycotoxins is based on a system of united rings called trichothecene (Godtfredsen et al., 1967). Natural trichothecenes contain a double bond at the C-9 - C-10 position and an epoxy group at the 12th and 13th carbon atoms. Trichothecene mycotoxins are highly chemically resistant and thermostable. Under natural conditions, they are practically unaffected by natural environmental factors. One is deoxynivalenol (DON, vomitoxin) (Bamburg, 1983). Researchers studied the effects of 12,13-epoxytrichothecenes (8-acetylneosalaniol, diacetoxyscirpenol, T-2 toxin, NT-2 toxin, neosalaniol, diacetyl-NT-2 toxin, and T-2 tetraol) on the organisms of day-old broiler chickens in Minnesota (Chi et al., 1979). They observed the appearance of symptoms such as appetite, asthenia, diarrhea, and coma in the experimental birds, which, in their opinion, indicated that the toxicity of mycotoxins depends on the modification of links in the structure of toxin molecules.

The scientist (Viczeko, 2019) proved that at the highest dose of DON (5.0 µg/g egg weight), it reduced the viability of chicken embryos and increased absolute indicators: the relative weight of the liver and spleen caused simultaneous bile stagnation in the liver and spleen inflammation. There was a dose-dependent increase in granulopoiesis and lipid peroxidation in the liver. However, the mRNA expression of genes associated with immune and oxidative stress in chicken embryos remained unchanged. His results indicate

that the chicken embryo responds to the introduction of DON by affecting its immunity and oxidative status. Mycotoxins in poultry feed are an important factor in financial losses in poultry production. DON can cause toxicological and immunotoxic effects in chickens. The main effects at low concentrations are reduced feed intake, while higher doses cause severe weight loss and impaired resistance to infection, particularly bacterial infection (Qu et al., 2019; Yao & Long, 2020; Sun et al., 2022; Hou et al., 2023). One crucial aspect of DON toxicity is damage to the gastrointestinal tract of animals. DON affects the digestive organs of chickens, especially the duodenum and cecum, as evidenced by their shorter and thinner villi. In addition, this toxin impairs intestinal function by reducing the absorption of nutrients (glucose and amino acids). There is evidence that DON impairs immune function in broiler chickens. Feeding grain contaminated with DON reduces serum antibody titers against Newcastle disease virus (NDV) and infectious bronchitis virus (IBV) in laying hens and broilers.

Researchers (Girish & Smith, 2008) found that mycotoxins negatively affect the intestinal barrier's function, reducing the intestinal epithelium's integrity. Apoptosis, increased colonization of pathogenic microorganisms, cytotoxicity, oxidative stress, inhibition of protein synthesis, and lipid peroxidation are characteristic of the toxic effects of mycotoxins on the intestinal epithelium. They directly or indirectly affect the host's immune responses (Brezvyn et al., 2021). Such immunotoxic effects of mycotoxins make poultry susceptible to many infectious diseases. The administration of silver deoxynivalenol at a dose of 70 mg/kg body weight to Adler cross chickens caused a decrease in body weight, changes in the activity of alkaline phosphatase isozymes and the metabolism of certain macronutrients in the blood serum, accompanied by pathological changes in the kidneys, heart, and liver (Ostrovskiy et al., 2014; Ostrovskiy, 2016).

At present, there is an urgent need to implement veterinary and sanitary, and preventive measures to develop and introduce into production new methods and means of prevention and treatment of mycotoxicosis in animals and poultry based on the use of natural or artificial sorbents together with toxin-contaminated feed.

The work aimed to study the quantitative and qualitative composition of microscopic fungi of wheat grain grown in the Polissya region of Ukraine and their toxigenic potential when fed to Adler silver sorbent "Mikosorb" to chickens.



2. Materials and methods

As a result of mycological studies, 53 samples of wheat grain from the Polissya region were used. Samples for re- search were collected from agricultural enterprises of vari- ous forms of ownership, the private sector, elevators, breed- ing stations, and regional seed inspections of Ukraine ac- cording to the guidelines for sanitary and mycological as- sessment and improvement of feed quality (Andrijchuk et al., 2011), following GOST 13586, 3-83 and DSTU 3570-

97. For in-depth studies of the toxin-synthesizing capacity of DON microbes, a wheat grain culture of var: "Remeslivna" from the Myronivsky Institute of Wheat named after

V. M. Remesla of the Ukrainian Academy of Agrarian Sci- ences and the Institute of Plant Physiology and Genetics of the National Academy of Sciences. This variety has been widely used in agricultural enterprises in all climatic zones of Ukraine (steppe, forest-steppe, and Polissya) since its state registration in 2004.

This wheat variety is a medium early variety with a pro- tein content of 14.3 %. The resistance to the lodging of this variety is 8–9 points, resistance to shattering – 8 points, resistance to root rot – 7–8 points, resistance to Septoria – 7–8 points, resistance to brown rust – 7–8 points, resistance to powdery mildew – 7–8 points. Since this variety is grown in different climatic conditions, we chose it as a generalized sample susceptible to micromycetes for further research.

The epiphytic mycorrhizal flora was studied by

$$C = \frac{Km}{V} \times N$$

$$N = \frac{N_1 + N_2 + \dots + N_m}{Km \left(\frac{1}{K_1} + \frac{1}{K_2} + \dots + \frac{1}{K_m} \right)}$$

3. Results and discussion

3.1. Results

During the research period, an average of $2.88 \cdot 10^4 \pm 3.62 \cdot 10^3$ colony forming units (CFU) per 1 g of grain was found in wheat grain samples collected in the Polissya re- gion. The results of the studies described in Table 1 show that the most commonly isolated fungi among the epiphytic mycobiota were *Mucor* spp. and *Alternaria alternata*. Less frequently in the studied wheat samples, we observed the infection with *Aspergillus fumigatus*, *Aspergillus flavus*, *Penicillium* spp., and *Fusarium sporotrichiella*. All other fungi we identified were found only in individual samples in isolated cases.

We identified 20 species of microscopic fungi

direct inoculation, for which wheat grains were placed in Petri dishes on the surface of Chapek's medium in 6–7 pieces. The material from each sample was sown in 4 Petri dishes, two cultivated at 24 °C and the other at 37 °C. Pure cultures were obtained by inoculating the fungi into tubes on Chapek's slant agar, and their cultural properties were taken into account, and microscopy was performed to determine the species. In order to determine the endophytic composi- tion of the mycobiota, the grain was treated with a 3 % formalin solution for 3 min before sowing, after which the material was washed with sterile distilled water to neutralize the disinfectant, to which a 5 % ammonia solution was add- ed.

The method of serial dilutions was used to determine the degree of contamination of the material with micromycetes (the number of colony-forming units per 1 g of grain). The grain was ground in an electric mill, and serial dilutions were prepared. In order to perform this, a 10 g sample of crushed grain was poured into 100 cm³ of sterile distilled water and shaken for 20 min, and serial dilutions of 1 : 100, 1 : 1000, and 1 : 10000 were prepared. Subsequently, 1 cm³ of the prepared dilutions was inoculated onto the surface of Chapek's medium at one dilution per two Petri dishes. Incu- bation was carried out at 24 and 37 °C, and colonies were counted on days 3 and 5 after sowing. The content of colo- ny-forming units per 1 g of wheat grain was calculated using the following formulas:

among the epiphytic mycobiota. Among mucoral fungi, *Absidia corymbifera* (32.1 %) and *Rhizopus oryzae* (24.5 %) were also detected in the samples.

Aspergillus niger (17.0 %), *Aspergillus candidus* (9.4 %), and *Aspergillus terreus* (1.9 %) were less common among the aspergilli. *Fusarium* spp. (17.0 %), *Fusarium oxysporum*, *Fusarium moniliforme*, *Fusarium semitectum* in (5.7 %), and *Fusarium culmorum* (3.8 %) of the samples were identified.

When performing mycological studies of endophytic mycobiota (Table 2), we often identified the following fun- gi: *Alternaria alternate*,



Aspergillus flavus, and *Mycelia sterilia*. In the selected wheat samples, grain damage by fungi of the genus *Mucor* spp: *Mucor* spp. *Penicillium* spp. *Mycelia sterilia* and *Phoma exiqua*. This indicates that fewerspecies of microscopic fungi represent the endophytic my- Where C is the content of fungal diaspores in 1 g of the substrate; K₁, K₂, Km, etc. are the denominators of the max- imum and used dilutions, starting with the minimum; V is the volume of the sown mixture (cm³); N₁, N₂, etc. is the number of diaspores, first the average, and then in each dilution (DSTU 8051:2015).

Identification of isolates to species and varieties was car- ried out using commonly accepted keywords (Domsch et al., 1980; Obrazhej et al., 1998; Ruhljada et al., 2009; Ostrovskij et al., 2018). The sizes of conidia, mycelium, and other elements of fungi were determined by crushed drop microscopy using an eyepiece micrometer.

The toxicity of the isolated isolates was determined by microbiological and mycotoxicological methods using thin- layer chromatography. To study the effect of DON on Adler silver chickens when fed the sorbent "Mycosorb" in 2 % of the total feed weight, the birds were divided into three groups of 100 heads each according to the principle of ana- logs. Chickens were kept in deep litter, each group in a sepa- rate section. Birds in the control group were fed a complete diet that did not contain toxins, while experimental group No. 1 was fed wheat grain containing the toxin deoxyniva- lenol as part of the feed ("Experimental (T)"). Experimental group No. 2 "(M+T)" (toxin + mycosorb) – received com- plete feed with wheat grain containing deoxynivalenol and mycosorb in the amount of 2 % by weight of the feed. The birds were constantly monitored, weighed weekly, and blood was taken for biochemical studies. The research re- sults were statistically analyzed using the built-in statistical functions of MS Excel and Stat Soft "Statistica 10" soft- ware. cobiota of the wheat grain.

Among the isolates of micromycetes isolated from the Polissya region, pure cultures were obtained

from *F. spo- rotrichiella* var. *tricinctum* isolate 1218/4, and *F. spo- rotrichiella* isolate 1218/5. Both isolates were atoxic against the test culture *Candida pseudotropicalis* strain 44 PC, but *F. sporotrichiella* var. *tricinctum* 1218/4 produced a growth retardation zone with Rf 0.05 and produced an unidentified trichothecene mycotoxin (TTMT). Among the isolated fun- gi, *Aspergillus flavus* isolate 1219/3 and *Aspergillus flavus* isolate 1221/1 were the first to produce kojic and aspergillic acids and the second to synthesize penicillic and aspergillic acids.

Microscopic fungi are permanent contaminants of grain crops. Under susceptible environmental conditions of tem- perature and humidity, they can synthesize secondary me- tabolites – mycotoxins. One of the most commonly detected mycotoxins in grain crops is deoxynivalenol (DON, vom- itoxin), produced by fungi of the genus *Fusarium*. In order to study the possibility of reducing the negative impact of the mycotoxin deoxynivalenol on the body of chickens of the meat and egg breed Adler Silver, the birds were fed the sorbent "Mikosorb". The study was conducted at a poultry enterprise under the conditions of the Educational and Sci- entific Research Center (ESRC) of Bila Tserkva NAU. As a result of the experiment (Table 3), it was found that when added to the diet, the sorbent "Mikosorb" in the amount of 2.0 % of the total feed weight reduces the nega- tive effect of deoxynivalenol on the body of chickens of the experimental group. This was confirmed by a 12.0 % reduc- tion in poultry mortality. Feeding "Microsorb" in the amount of 2.0 % by weight of complete feed contributed to an increase in the average daily weight gain of poultry dur- ing the experiment by 5.43 % compared to the experimental group that consumed feed with DON toxin. During the ex- periment, the birds of the experimental group No. 2 con- sumed 28,91 kg of feed more than group No. 2. Feeding of "Mikosorb" to chickens of experimental group 2 contributed to an increase in the level of profitability of poultry rearing up to 12.0 % compared to poultry that consumed feed af- fected by DON toxin.

Table 1

Epiphytic mycobiota of wheat grain in the Polissya zone

Types of isolated micromycetes	Polissya region (53 samples)	
	Fact. (number of samples)	% of identified
<i>Zygomycota, Zygomycetes, Mucorales, Mucoraceae</i>		
<i>Mucor</i> spp.	49	92.5
<i>Absidia corymbifera</i> (Cohn) Jacc. Et A. Trotter	17	32.1
<i>Rhizopus oryzae</i> Went.	13	24.5
Together mucoral	44	83
<i>Ascomycota, Plectomycetes, Eurotiales</i>		
<i>Monascus rubber</i> van Tieghem	1	1.9
<i>Mitosporic fungi, Coelomycetes, Sphaeropsidales, Sphaeroidaceae</i>		
<i>Phoma exiqua</i> Desmazieres	8	15.1
<i>Hyphomycetales, Dematiaceae</i>		



<i>Alternaria alternata</i> (Fr.) Keissl	49	92.5
<i>Moniliaceae</i>		
<i>Aspergillus fumigatus</i> Fres.	31	58.5
<i>Aspergillus flavus</i> Zr:Fr	28	52.8
<i>Aspergillus niger</i> van Tieghem	9	17
<i>Aspergillus candidus</i> Zk:Fr.	5	9.4
<i>Aspergillus terreus</i> Thom.	1	1.9
Together aspergillus	44	83
<i>Penicillium</i> spp.	25	47.2
<i>Trichothecium roseum</i> (Pers:Fr.) Zk.	1	1.9
<i>Hyphomycetales, Agonomycetales, Agonomycetaceae</i>		
<i>Mycelia sterilia</i> (Pink)	8	15.1
<i>Tuberculariales, Tuberculariaceae</i>		
<i>Fusarium</i> spp.	9	17
<i>Fusarium sporotrichiella</i> Bilai	16	30.2
<i>Fusarium oxysporum</i> (Schlecht.) Snyd. Et. Hans	3	5.7
<i>Fusarium moniliforme</i> Sheld.	3	5.7
<i>Fusarium semitectum</i> Berk. et. Rav	3	5.7
<i>Fusarium culmorum</i> (W.G.Sm.) Sacc.	2	3.8
Together fusariums	32	60.4

Table 2

Endophytic mycobiota of wheat grain in the Polissya zone

Types of isolated micromycetes	Polissya region (53 samples)	
	Fact. (number of samples)	% of identified
<i>Zygomycota, Zygomycetes, Mucorales, Mucoraceae</i>		
<i>Mucor</i> spp.	11	20.8
Together mucoral	11	20.8
<i>Mitosporic fungi, Coelomycetes, Sphaeropsidales, Sphaerioidaceae</i>		
<i>Phoma exiqa</i> Desmazieres	9	17
<i>Hyphomycetales, Dematiaceae</i>		
<i>Alternaria alternata</i> (Fr.) Keissl	52	98.1
<i>Moniliaceae</i>		
<i>Aspergillus fumigatus</i> Fres.	3	5.7
<i>Aspergillus flavus</i> Zr:Fr	22	41.5
<i>Aspergillus niger</i> van Tieghem	1	1.9
Together Aspergillus	25	47.2
<i>Penicillium</i> spp.	11	20.8
<i>Trichothecium roseum</i> (Pers: Fr.) Zk.	1	1.9
<i>Hyphomycetales, Agonomycetales, Agonomycetaceae</i>		
<i>Mycelia sterilia</i> (Pink)	12	22.6
<i>Tuberculariales, Tuberculariaceae</i>		
<i>Fusarium</i> spp.	3	5.7
<i>Fusarium sporotrichiella</i> Bilai	3	5.7
<i>Fusarium oxysporum</i> (Schlecht.) Snyd. Et. Hans	5	9.4
<i>Fusarium moniliforme</i> Sheld.	3	5.7
Together fusariums	13	24.5

Table 3

Economic efficiency of feeding Mycosorb to Adler Silver chickens (M ± m, n = 100)

Indicators	Groups of poultry		
	Control	Experimental № 1 (T)	Experimental No. 2 (M+T)
The initial number of chickens, goal.	100	100	100
Number of livestock at the end of the experiment, goal.	91	74	86
Preservation, %.	91.0	74.0	86.0
The average number of livestock during the study period, heads.	95.5	87.0	93.0
Total weight of birds at the beginning of the experiment (4 weeks old) kg	31.81 ± 0.133	31.57 ± 0.129	31.35 ± 0.125
Average live weight of 1 head at the beginning of the experiment (age four weeks), g	318.1 ± 6.11	315.7 ± 7.19	313.5 ± 8.17
Total weight of poultry at the end of the experiment (7 weeks of age) kg	53.15 ± 0.143	42.00 ± 0.121	51.47 ± 0.132
Average live weight of 1 head at the end of the experiment (age seven weeks) g	584.1 ± 11.21	567.7 ± 14.19	598.5 ± 12.16
Feed consumption during the experiment (4-7 weeks), kg	86.24 ± 0.381	51.16 ± 0.279	80.07 ± 0.374



Cost of feed during the experiment (4-7 weeks), UAH	238.02	141.20	220.99
The cost of using Mycosorb during the experiment (4-7 weeks of 2% to feed), UAH	-	-	138.36
Electricity cost for the period of the experiment, UAH	197.26	197.26	197.26
The cost of dead poultry during the growing process, UAH	145.80	421.20	226.80
Other production costs, UAH	172.25	172.25	172.25
Gross live weight gain obtained during the experiment, kg	21.34 ± 0.229	10.43 ± 0.193	20.12 ± 0.215
The cost of raising poultry for the period of the experiment, UAH	753.33	931.91	800.66
Cost of live weight gain in poultry, UAH	950.70	464.86	896.74
Profit from live weight gain, for the period of research, UAH	197.37	-467.05	96.08
Profitability, %	26.2	-	12.0

3.1. Discussion

The contamination of wheat grain with micromycetes largely depends on the region's climatic conditions, the cultivation period, and compliance with storage conditions (Vasjanovych et al., 2016). Our results indicate that the Polissya zone is characterized by higher precipitation and, accordingly, higher humidity, contributing to increased wheat grain contamination with microscopic fungi. The increase in the number of isolated species of microscopic fungi causes the

detection of a significant variety and concentration of mycotoxins in the grain. A similar positive correlation between the number of fungal species and their toxicity was noted by other authors (Vasjanovych et al., 2017). The introduction of 2.0 % by weight of feed increased bird productivity by 5.43 %. A positive effect on increasing the productivity and safety of chickens with the use of sorbents in bird feeding was reported by other authors (Kotsyumbas et al., 2010).

4. Conclusions

Twenty microscopic fungi from 9 genera were isolated from wheat grain in the Polissya region. Among them were the genera *Alternaria* (92.5 %), *Mucor* (92.5 %), *Aspergillus* (83.0 %), *Penicillium* (47.2 %), *Fusarium* (60.4 %), *Phoma* (15.1 %), *Mycelia* (15.1 %), *Trichotecium* (1.9 %) and *Monascus* (1.9 %) of the samples. One of them was a producer of trichothecene mycotoxin; the other two synthesized kojic, penicillic and aspergillic acids.

Introducing the silver sorbent "Mikosorb" in the amount of 2.0 % by weight of feed into the composition of mixed fodder for chickens of the

meat and egg breed Adler contributed to an increase in bird productivity by 5.43 %. Using the sorbent "Mikosorb" increases the safety of meat and egg breed Adler silver chickens by 12.0 %. Feeding the sorbent "Mikosorb" in an amount of 2.0 % of the feed weight to chickens of the meat and egg breed Adler Silver contributes to an increase in gross weight gain by 9.69 kg during the experiment. After analyzing the scientific results from the literature and our research results, we concluded that the above-mentioned studies on wheat grain damage should be conducted throughout Ukraine during harvesting and storage in warehouses or storages.

References

- Andrijchuk, A. V., Bilan, A. V., Sydorchuk, P. I., & Ostrovs'kyj, D. M. (2011). Toksygenni vlastyvoli mikromicetiv zerna pshenyca ta jachmenju. *Visnyk agrarnoi nauky*, 9, 22–24 (in Ukrainian). [\[Article\]](#)
- Antonjak, G. L., Fedjakov, R., Koval', N., & Stefanyshyn, O. (2010). Vplyv mikotoksyniv na zdorov'ja tvaryn. *Naukovyj visnyk veterynarnoi medycyny*, 5(78), 10–13 (in Ukrainian). [\[Article\]](#) [\[Google Scholar\]](#)
- Bamburg, J. R. (1983). Biological and biochemical actions of trichothecene mycotoxins. *Progress in molecular and subcellular biology*, 8, 41–110. [\[Article\]](#) [\[Google Scholar\]](#)
- Beccari, G., Prodi, A., Senatore, M. T., Balmas, V., Tini, F., Onofri, A., & Covarelli, L. (2020). Cultivation area affects the presence of fungal communities and secondary metabolites in Italian durum wheat grains. *Toxins*, 12(2), 97. [\[Crossref\]](#) [\[Google Scholar\]](#)
- Brezvyn, O. M., Guta, Z. A., Gutyj, B. V., Fijalovych, L. M., Kar-povskiy, V. I., Shnaider, V. L., Farionik, T. V., Dankovych, R. S., Lisovska, T. O., Bushuieva, I. V., Parchenko, V. V., Magre-lo, N. V., Slobodjuk, N. M., Demus, N. V., & Leskiv, Kh. Ya. (2021). The influence of HamekoTox on the morphological and biochemical indices of the blood of laying hens in spontaneous fumonisin toxicosis. *Ukrainian Journal of Ecology*, 11(2), 249–253. [\[Article\]](#) [\[Google Scholar\]](#)
- Chi, M. S., Robison, T. S., Mirocha, C. J., & Reddy, K. R. (1978). Acute toxicity of 12, 13-epoxytrichothecenes in one-day-old broiler chicks. *Applied and Environmental Microbiology*, 35(4), 636–640. [\[Crossref\]](#) [\[Google Scholar\]](#)
- Domsch, K. H., Gams, W., & Anderson, T. H. (1980). *Compendium of soil fungi. Volume 1*. Academic Press (London).



- [[Google Scholar](#)]
DSTU 8051:2015 (2015). *Produkty kharchovi*. Metody vidbyran-nia prob dlia mikrobiolohichnykh analiziv (in Ukrainian). [[DSTU](#)]
- Dvorska, Ju. Je. (2013). Mikotoksyny v kormah ptyci: ocinka ryzyku. *Agrarnyj visnyk Prychornomor'ja. Veterynarni nauky*, 68, 62–69. (in Ukrainian). [[Article](#)] [[Google Scholar](#)]
- Felšöciová, S., Kowalczewski, P.L., Krajčovič, T., Dráb, Š., & Kačaniová, M. (2021). Effect of Long-Term Storage on Myco-biota of Barley Grain and Malt. *Plants (Basel)*, 10(8), 1655. [[Crossref](#)] [[Google Scholar](#)]
- Girish, C., & Smith, T. (2008). Impact of feed-borne mycotoxins on avian cell-mediated and humoral immune responses. *World Mycotoxin Journal*, 1(2), 105–121. [[Crossref](#)] [[Google Scholar](#)]
- Godtfredsen, W. O., Grove, J. F., & Tamm, C. (1967). On the nomenclature of a new class of sesquiterpens. *Helvetica Chim-ica Acta*, 50, 1666–1668. [[Google Scholar](#)]
- Golovchak, N. (2007). Struktura ta vplyv mikotoksyniv na zhyvi organizmy. *Visnyk Lvivskoho universytetu. Seriya biolohichna*, 43, 33–47 (in Ukrainian). [[Google Scholar](#)]
- Hmel'nyckyj, G. O., Malinin, O. O., Kucan, O. T., & Duhnyckyj, V. B. (2012). *Veterynarna toksykologija*. Kyiv: Agrarna osvita (in Ukrainian). [[Google Scholar](#)]
- Hou, S., Ma, J., Cheng, Y., Wang, H., Sun, J., & Yan, Y. (2023). The toxicity mechanisms of DON to humans and animals and potential biological treatment strategies. *Critical Reviews in Food Science and Nutrition*, 63(6), 790–812. [[Crossref](#)] [[Google Scholar](#)]
- Illif, G. J., Mukherjee, R., Gruszewski, H. A., Schmale Iii D. G., Jung, S., & Boreyko, J. B. (2022). Phase-change-mediated transport and agglomeration of fungal spores on wheat awns. *Journal of The Royal Society Interface*, 19(190), 20210872. [[Crossref](#)] [[Google Scholar](#)]
- Jangol, Ju. A. (2018). Vyznachennja toksychnosti ta toksynout-vorennja mikroskopichnyh grybiv v kormah. *Veterynarna bio-tehnologija*, 33, 130–135 (in Ukrainian). [[Crossref](#)] [[Google Scholar](#)]
- Jaroshenko, M. O., Kucan, O. T., & Orobchenko, O. L. (2018). Monitoryng kormiv dlja VRH molochnogo naprjamu produktyvnosti na najavnist' plisenevyh mikromicetiv u gospodarstvah pivnichno-shidnogo regionu Ukrainy. *Veterynarna bio-tehnologija*, 32(2), 602–610 (in Ukrainian). [[Crossref](#)] [[Google Scholar](#)]
- Kolomiets, T. M., Kiseleva, M. I., Zhemchuzhina, N. S., Pankratova, L. F., & Elizarova, S. A. (2022). A characteristic of the species composition of pathogenic fungi of the genus *Fusarium* in corn biocenoses of the Voronezh region. *Vavilovskii Zhurnal Genet Seleksii*, 26(6), 583–592. [[Crossref](#)] [[Google Scholar](#)]
- Kotsyumbas, I. Ja., Avdos'jeva, I., Brezvyin, O., Temnenko, S., Regenchuk, V., & Basarab, O. (2010). Efektyvnist' vakcyvacii' proty virusnyh zahvorjuvan' ptyci u razi zastosuvannja detoksykantiv mikotoksyniv. *Naukovyi visnyk veterynarnoi medytsyny*, 63, 63–69 (in Ukrainian). [[Article](#)] [[Google Scholar](#)]
- Kotyk, A. M., Trufanova, V. O., Gorbenko, Z. G., & Chorna, G. V. (2013). Dija nyz'kykh koncentracij zearalenonu v kormi na pivniv. *Ptakhivnytstvo*, 69, 149–156 (in Ukrainian). [[Article](#)]
- Minooecianhaghghi, M. H., Marvi Moghadam Shahri, A., & Taghavi, M. (2021). Investigation of feedstuff contaminated with aflatoxigenic fungi species in the semi-arid region in northeast of Iran. *Environmental Monitoring and Assessment*, 193(4), 214. [[Crossref](#)] [[Google Scholar](#)]
- Obrazhej, A. F., Pogrebnjak, L. Y., & Korzunenko, O. F. (1998). Metodychni vkazivky po sanitarnomikologichnij ocinci ta polipshennju jakosti kormiv: Zatverdzh Derzhavnym departamentom veterynarnoi medycyny Ministerstva APK Ukrainy vid 06.03. 1998. № 15-14/73. Kyi'v (in Ukrainian). [[Google Scholar](#)]
- Ostrovskij, D. M. (2016). Biohimichni zminy syrovatky krovi kurchat vnaslidok dii' dezoksynivalenolu. *The Animal Biology*, 18(3), 71–77 (in Ukrainian). [[Crossref](#)] [[Google Scholar](#)]
- Ostrovskij, D. M., Andrijchuk, A. V., & Zocenko, V. M. (2018). Mikromicety zerna pshenyци v Ukraini. *Naukovyj visnyk veterynarnoi medycyny*, 1(140), 116–122 (in Ukrainian). [[Article](#)] [[Google Scholar](#)]
- Ostrovskij, D. M., Mel'nyk, A. Ju., & Utechenko, M. V. (2014). Vyvchennja vplyvu dezoksynivalenolu na kurchat krosu Adler sribljastyj ta profilaktychnoi' dii' mikosorbu. *Naukovyj visnyk veterynarnoi medycyny*, 14, 151–156 (in Ukrainian). [[Crossref](#)] [[Google Scholar](#)]
- Qu, R., Jiang, C., Wu, W., Pang, B., Lei, S., Lian, Z., Shao, D., Jin, M., & Shi, J. (2019). Conversion of DON to 3-epi-DON in vitro and toxicity reduction of DON in vivo by *Lactobacillus rhamnosus*. *Food & Function*, 10(5), 2785–2796. [[Crossref](#)] [[Google Scholar](#)]
- Rozhkova, T. O., Tatarynova, V. I., & Burdulanjuk, A. O. (2017). Osoblyvosti identyfikacii' vydiv endofitnoi' mikrobioty nasinnjapshenyци ozymoi' z pivnichnogo shodu Ukrainy. *Visnyk Sums'kogo nacional'nogo agrarnogo universytetu. Serija: Agronomija i biologija*, 9, 6–12 (in Ukrainian). [[Article](#)] [[Google Scholar](#)]
- Ruhljada, V. V., Ostrovskij, D. M., & Kurchenko, I. M. (2009). Epifitna i endofitna mikrobiota zerna



pshenyaci v Ukraini. *Nau- kovyj visnyk veterynarnoi' medycyny*, 62, 80-83 (in Ukrainian). [[Google Scholar](#)]

Stuper-Szablewska, K., & Perkowski, J. (2014). Contamination of wheat grain with microscopic fungi and their metabolites in Poland in 2006-2009. *Annals of Agricultural and Environmental Medicine*, 21(3), 504-509. [[Crossref](#)] [[Google Scholar](#)]